The Structure of Baccatin-III, a Partially Esterified Octahydroxy-monoketo-taxane Derivative Lacking a Double Bond at C-4

By D. P. DELLA CASA DE MARCANO, T. G. HALSALL,* and G. M. HORNBY (The Dyson Perrins Laboratory, Oxford University, Oxford, OX1 3QY)

Summary Baccatin-III is the triacetate monobenzoate of 1β , 2α , $4(?\beta)$, 5α , 7β , 10β , 13α , 20-octahydroxytax-11-en-9-one.

RECENTLY we described¹ the isolation of a complex ester. baccatin-III, from the heartwood of yew (Taxus baccata L.) and provisionally formulated it as C₃₁H₃₈O₁₁. It has now been shown to be $C_{33}H_{42}O_{13}$ (containing the elements of one more acetic acid than the original formulation) and to have m.p. 233–234°, $[\alpha]_{D} - 93^{\circ}$, λ_{max} 230, 274, and 280 nm, ϵ 13,900, 1000, and 850. Structure (I) is suggested for the ester. It is thus the first example of a naturally occurring taxane derivative which lacks the exocyclic double bond at C-4. The n.m.r. spectrum of baccatin-III showed the signals of the four methyl groups for a taxane system at τ 7.75, 8.35, 8.91, and 8.91. The presence of three acetate and one benzoate groups was shown by spectroscopic data. An n.m.r. signal at τ 3.66 (s, 1H) and the absence of the pair of doublets associated with the 9,10-dioxy-grouping, together with the large negative rotation and the position of the bands due to the methyl groups, indicated a 9-oxo- 10β -acetoxy-grouping analogous to that found in the ketol acetates (II)² and (III)³. The presence of two secondary hydroxy-groups was shown by n.m.r., and one of them was readily acetylated to give baccatin-III monoacetate (IV). Two tertiary hydroxygroups accounted for the remaining oxygen atoms. No signals, however, were found due to two olefinic protons at C-20. Instead the presence of an AB system at τ 5.70 and 5.83 (J 8 Hz) suggested the presence of a C-CH₂-O·CO·Ph grouping. A doublet at $\tau 4.38$ (J 7 Hz) was found to be coupled with a doublet at τ 6.14. This latter signal was assigned to the C-3 proton which indicated that the signal at τ 4.38 was due to a proton on C-2 which carried an α -acetoxy-group. It also follows that there is no proton at C-1, which must be substituted with a tertiary hydroxygroup. The low τ value of the C-3 proton supported the presence of a 9-oxo-group which causes deshielding of this proton similar to that observed for the compound (II).4

When baccatin-III monoacetate (IV) was hydrogenated with Adams catalyst in acetic acid, the n.m.r. spectrum of the product (V) showed the absence of a signal for one

proton at τ 5.15 indicating the loss of one allylic secondary hydroxy-group from C-13. This was confirmed by prolongated treatment of baccatin-III with sodium metaperiodate when oxidation of the allylic hydroxy-group occurred with the formation of an $\alpha\beta$ -unsaturated ketone (VI) (λ_{max} 275 nm, ϵ 6800) which showed the abnormal light adsorption associated with the 13-oxo-11-ene system of the taxanes. Although no fission of a glycol group occurred under these conditions, baccatin-III gave a positive result for an α-glycol group with Fiegel's periodic acid oxidation test.⁵ Since the C-3 proton is only coupled to one proton at C-2, there must be a hydroxy-group at C-4 and this group together with a secondary hydroxy-group at C-5 would account for the α -glycol structure. The C-5 hydroxygroup was the one which was acetylated, and when this took place the τ signal for the C-5 proton shifted from au 5.54 (q, J 10.5, 6 Hz) to au 4.36 (q, J 10, 6 Hz). The remaining acetoxy-group is placed at C-7, the proton geminal with it having a signal at τ 5.01 (q, J 10, 4 Hz).

If the substituents in baccatin-III have the same stereochemistry as in other taxane derivatives² the C-7 acetoxygroup should be β and the C-5 hydroxy-group α . The coupling constant of the proton at C-7 is consistent with the values given for other taxane derivatives,^{6,7} but that of the proton at C-5 is different from that usually found, and indicated that it is axial rather than equatorial. Inspection of models indicates, however, that with a 5α -hydroxy-group there is a favourable conformation of baccatin-III with ring c present as a boat with C-5 and C-8 as the bow and stern, and this conformation gives dihedral angles which account for the coupling constants for both the 5 β - and 7 α protons. Finally there is the question of the stereo-chemistry at C-4. The most likely biogenetic route to the C-4, C-20 dioxy-system is via epoxidation of the C-4, C-20

double bond and opening of the oxide ring. If so the α -epoxide is the more likely to be formed. Assuming *trans*-opening of the epoxy-group, this would lead to 4β hydroxy-group. This stereochemistry would give a trans- α -glycol grouping in baccatin-III which could account for the resistance to oxidation with sodium metaperiodate.



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